Contents lists available at ScienceDirect

Cancer Treatment Reviews

journal homepage: www.elsevierhealth.com/journals/ctrv

Therapeutic implications of resistance to molecular therapies in metastatic colorectal cancer

A. Sartore-Bianchia, *, K. Bencardino^a, A. Cassingena^a, F. Venturinia, C. Funaiolia, T. Cipania, A. Amatua, L. Pietrogiovanna^a, R. Schiavo^a, F. Di Nicolantonio^{b, c}, S. Artale^a, A. Bardelli^{b, c}, S. Siena^a

^a The Falck Division of Medical Oncology, Department of Oncology, Ospedale Niguarda Ca' Granda, Milano, Italy

^b Laboratory of Molecular Genetics, Institute for Cancer Research and Treatment (IRCC), University of Torino Medical School, Candiolo, Turin, Italy ^c FIRC, Institute of Molecular Oncology, Milan, Italy

article info

Keywords: Colorectal cancer *KRAS* EGFR Monoclonal antibodies

SUMMARY

Metastatic colorectal cancer (mCRC) patients carrying KRAS mutated tumors do not benefit from epidermal growth factor receptor (EGFR)-targeted cetuximab- or panitumumab-based therapies. Indeed, the mutational status of *KRAS* is currently a validated predictive biomarker employed to select mCRC patients for EGFR targeted drugs. When patients fail standard 5-fluorouracil-, oxaliplatin-, irinotecan- and bevacizumab-based therapies, EGFR-targeted salvage therapy can be prescribed only for those individuals with *KRAS* wild-type cancer. Thus, clinicians are now facing the urgent issue of better understanding the biology of *KRAS* mutant disease, in order to devise novel effective therapies in such defined genetic setting. In addition to *KRAS*, recent data point out that *BRAF* and *PIK3CA* exon 20 mutations hamper response to EGFR-targeted treatment in mCRC, potentially excluding from treatment also patients with these molecular alterations in their tumor. This review will focus on current knowledge regarding the molecular landscape of mCRC including and beyond KRAS, and will summarize novel rationally-developed combinatorial regimens that are being evaluated in early clinical trials.

© 2010 Elsevier Ltd. All rights reserved.

Background

The introduction of *KRAS* testing as a diagnostic tool to select patients for epidermal growth factor receptor (EGFR)-targeted cetuximab- or panitumumab-based therapies for metastatic colorectal cancer (mCRC) has been validated and is regarded as one of the most important recent advances in the field of personalized cancer medicine. ¹ The oncogene *KRAS* is indeed the most commonly mutated gene in various human cancers and it has been demonstrated that its constitutive activation in mCRC can bypass the EGFR-driven signalling cascade and impair the clinical efficacy of EGFR-targeted monoclonal antibodies. ² *KRAS* testing has rapidly led to an improvement in the therapeutic index of these drugs, excluding from treatment patients harbouring mutations in the tumor, who do not achieve clinical benefit from these targeted therapeutics, as recommended by the American Society of Clinical Oncology (ASCO). 3 Thus, clinicians are now facing the emerging issue of better understanding the biology of the *KRAS* mutant disease, because the unfeasibility of EGFR-targeted salvage therapy leaves an unmet need for treatment options in those patients

who fail standard 5-fluorouracil-, oxaliplatin-, irinotecan- and bevacizumab-based therapies.

In a recent paper by De Roock et al.,⁴ 649 tumour DNA samples from chemotherapy-refractory mCRC patients treated with cetuximab plus chemotherapy were gathered from 11 centres in seven European countries, and investigators show that also *BRAF, NRAS*, and *PIK3CA* exon 20 mutations were significantly associated with a low response rate, confirming findings from previous patients' cohort analyses.^{5,6} This suggests that also patients harbouring these molecular alterations should be excluded from EGFR-targeted treatment with cetuximab, thus lacking effective third-line treatment strategies. At present, each of these markers (*KRAS, BRAF, PIK3CA*) has been mainly assessed as a single event, often in retrospective analyses and patients series, but these molecular alterations display overlapping pattern of occurrence, thus adding complexity for drawing an algorithm suitable for clinical decision-making. For this reason, last-generation studies by our group and others nowadays include comprehensive integrated analysis of the entire oncogenic pathway triggered by the EGFR, with the aim of enhancing the prediction ability of the markers individually used. ⁷

There is an urgent need to develop effective salvage therapies for patients with primary refractoriness to EGFR targeted monoclonal antibodies (i.e. those affected by *KRAS*/*BRAF*/*PIK3CA* mutated tumors), as well for those who develop resistance over prolonged

^{*} Corresponding author. Andrea Sartore-Bianchi M.D. Divisione di Oncologia Falck, Ospedale Niguarda Ca' Granda, Piazza Ospedale Maggiore, 3, 20162 Milan, Italy.

E-mail address: andrea.sartorebianchi@ospedaleniguarda.it (A. Sartore-Bianchi).

treatment. This review will focus on agents targeting the KRAS pathway, given its pivotal role in the biology of CRC and response to anti-EGFR therapies. Additionally, we will outline clinical trials with drugs directed at molecular targets that might be synthetically lethal with concomitant EGFR inhibition.

Targeting molecular effectors downstream of EGFR interacting with RAS that could preclude responsiveness to cetuximab or panitumumab

BRAF

BRAF is a cytoplasmatic serine/threonine kinase directly interacting with RAS, which regulates its activity starting a cytoplasmic phosphorylation cascade which leads to the activation of transcription factors controlling cell growth, differentiation and apoptosis (the ERK signalling pathway). ⁸ The *BRAFV600E* mutation, a thymine to adenine transversion mutation, resulting in the substitution of valine with glutamate, appears in $4-15\%$ of CRC.^{4,7} Importantly, *KRAS* and *BRAF* mutations are known to be mutually exclusive in CRC,⁹ and, as for *KRAS*, there is a high concordance of *BRAF* mutations in primary CRC and related metastatic sites. ¹⁰ The first study assessing the role of the *BRAFV600E* mutation as a predictive molecular factor to EGFR-targeted therapy was performed by our group in a cohort of 113 tumors from patients who received panitumumab/cetuximab in second or subsequent lines of treatment.⁵ Results from this retrospective analysis showed that, among *KRAS* wild-type patients, those whose tumors displayed the *BRAFV600E* mutation (14%) did not respond to EGFR inhibition and had statistically significantly shorter progression-free survival (PFS) and overall survival than patients whose tumors carried wild-type *BRAF*. In the same article, we also demonstrated that introduction of the *BRAFV600E* allele could confer resistance to either cetuximab or panitumumab in wild-type *BRAF* CRC cells. Subsequently, Loupakis et al. performed a retrospective analysis among 87 irinotecan refractory patients, treated with anti-EGFR therapy.¹¹ They found that *BRAF* was mutated in 13 cases (15%): none of the patients bearing *BRAF* mutation responded to the treatment, in comparison with 24 (32%) of 74 patients with *BRAF* wild-type disease (p = 0.016). *BRAF* mutation was also associated with a trend towards shorter *PFS* and with significantly shorter overall survival. In the recent wide retrospective cohort analysis of chemorefractory patients from a European Consortium (n = 761), De Roock et al. reported 4.7% *BRAF* mutations: among *KRAS* wild-type patients treated with cetuximab a significantly lower response rate, shorter PFS and overall survival than with wild-type tumors were observed, and this effect was confirmed in multivariate analyses performed using the mutation status of *KRAS, PIK3CA* exon 20, *PIK3CA* exon 9, *BRAF*, and *NRAS*, and age, sex, number of previous chemotherapy lines, and European centre as covariates. ⁴

In summary, data from these^{4,5,11} and other studies⁷ clearly show that, in the CRC chemorefractory setting, *BRAF* mutations are predictive of resistance to EGFR-targeted monoclonal antibodies. Since RAF is an important effector downstream of RAS in the ERK signalling pathway, targeting this effector could be also viewed as an effective strategy for treating *KRAS* or *BRAF* mutated tumors. We showed indeed that one of the first inhibitors of RAF activity, sorafenib, may restore sensitivity to EGFR inhibitors in *BRAF*mutated CRC cell lines⁵ and, consequently, combined sorafenib and cetuximab therapy is undergoing clinical evaluation in mCRC in a National Cancer Institute-sponsored trial (ClinicalTrials.gov Identifier: NCT00343772, Table 1). Nevertheless, it should be taken into account that sorafenib is a multikinase inhibitor which does not work primarily as a RAF inhibitor, but rather as an inhibitor of angiogenesis by inhibiting VEGFR-1, -2, -3, and PDGFR, ¹² and thus better results would have been expected by more

selective RAF inhibitors such as PLX4032, that showed pronounced activity in *BRAF*-mutant melanoma patients. Unexpectedly, in a recent report from Kopetz et al., evaluating 19 mCRC *BRAFV600E* patients treated with PLX4032, only a modest clinical activity was observed (1 confirmed partial response, 4 minor responses with ≥10% shrinkage and 5 mixed responses), suggesting that the biology of BRAF activation in patients with mCRC is clearly more heterogeneous than in melanoma, as evidenced in those patients with a mixed response. ¹³ Interestingly, Hatzivassiliou et al. recently demonstrated that ATP-competitive RAF inhibitors, such as GDC-0879 and PLX4720, possess two opposing mechanisms of action depending on the cellular context, i.e., in *BRAFV600E* tumours, RAF inhibitors effectively block the MAPK signalling pathway and decrease tumour growth, while in *KRAS* mutant and *KRAS/BRAF* wild-type tumours, RAF inhibitors activate the RAF-MEK-ERK pathway in a RAS-dependent manner, thus enhancing tumour growth in xenograft models. ¹⁴ Conversely, the MEK inhibitor PD0325901 inhibits proliferation of *BRAFV600E* , *KRAS/BRAF* wild-type and *KRAS* mutant cancer cells. ¹⁴ Similar findings come also from the study by Poulikakos et al., showing that ATP-competitive RAF inhibitors inhibit ERK signalling in cells with mutant *BRAF*, but paradoxically enhance signalling in cells with wild-type *BRAF* by drug-mediated transactivation of RAF dimers. ¹⁵

From a clinical standpoint, these preclinical results indicate the working hypothesis that RAF inhibitors may be used in mCRC in which *BRAF* only is mutated, whereas MEK inhibitors could be effective in a wider range of conditions: *BRAFV600E* , *KRAS/BRAF* wildtype and *KRAS* mutant tumors. It remains to be shown whether concomitant blockade of EGFR and BRAF or MEK would result in increased clinical efficay in BRAF/KRAS mutant tumors. Table 1 shows selected ongoing studies with EGFR-directed monoclonal antibodies in combination with other targeted agents, including BRAF inhibitors.

PIK3CA and PTEN

In addition to RAS and RAF, the EGFR also activates the PI3K signaling pathway, which in turn can be oncogenically deregulated either by activating mutations in the *PIK3CA* p110 subunit or by inactivation of the PTEN phosphatase. Importantly, there is an interaction between RAS and PI3K, since the PI3K signaling pathway can be activated both by EGFR as well as by RAS itself. ¹⁶ The role of deregulated *PIK3CA*/PTEN signaling on the response to targeted therapy has been investigated in breast, glioblastoma and also mCRC. *PIK3CA* mutations occur in approximately 10–18% of CRC patients, principally located in exon 9 and $20,4,6,17$ whereas loss of PTEN expression by immunohistochemistry (IHC) is reported in 19–42%. 18,19 *In vitro* studies in various CRC cell lines have found that activating *PIK3CA* mutations or loss of PTEN expression appear to confer resistance to cetuximab: cell lines carrying mutations in *PIK3CA*, or displaying loss of PTEN, with concomitant mutations in *RAS* or *BRAF* exhibit the greatest resistance to cetuximab. ²⁰ In the clinical setting, we found that in a cohort of 110 patients *PIK3CA* mutations and PTEN loss were statistically significantly associated with lack of response to panitumumab (0/15 patients, $p = 0.038$) or cetuximab (1/32 patients, $p = 0.001$) treatment.⁶ In the same study, *PIK3CA* mutations and/or loss of PTEN expression were negatively associated with PFS, and loss of PTEN expression was also linked with poorer overall survival ($p = 0.005$). This negative association with PFS was also noted in a study by Souglakos et al., ²¹ where among 92 patients treated using chemotherapy and cetuximab as salvage therapy, *PIK3CA* mutations predicted reduced PFS (2.5 vs 3.9 months, HR 2.1, 95% CI 1.2–3.9). In contrast, Prenen et al. reported in a series of 200 mCRC patients that 23 (12%) carried a *PIK3CA* mutation and 5 of these (22%) were found in responders. ²² This means that 5 of 39 responders (13%) and 18

Table 1

Selected clinical trials evaluating the combination of EGFR-directed therapeutics with other targeted agents. Trials were retrieved from the U.S. National Institutes of Health service *ClinicalTrials.gov* and updated as of July, 30th

NCT: ClinicalTrials.gov Identifier; mCRC: metastatic colorectal cancer.

of 160 non-responders (11%) carried a *PIK3CA* mutation, thus not supporting a significant association between *PIK3CA* mutations and lack of response to cetuximab ($p = 0.781$). The median PFS and overall survival did also not differ significantly between *PIK3CA* mutant and wild-type patients. Finally, the large dataset by the European Consortium showed that among 356 *KRAS* wild-type chemorefractory tumors treated with cetuximab, patients with mutant *PIK3CA* as a whole had a significantly lower response rate compared with carriers of wild-type *PIK3CA*, (17.7% [6/34] *vs* 37.7% [115/305]; OR 0.35, 95% CI 0.13–0.83; p = 0.015). Notably, there was no significant difference in PFS and overall survival (median PFS 18 vs 24 weeks, HR 1.30, 95% CI 0.91-1.86; p=0.17; and median overall survival 39 vs 51 weeks; HR 1.41, 0.96–2.06; p = 0.09). ⁴ However, when compared with *PIK3CA* wild-type, *PIK3CA* exon 20 mutations had a negative effect on objective response (0.0% [0/9] vs 36.8% [121/329], Fisher's exact test estimated OR 0.00, 95% CI 0.00–0.89; p = 0.029), disease control (33.3% [3/9] vs 76.0% [250/329]; OR 0.158, 0.0327–0.613; p = 0.0078), PFS (median 11.5 vs 24 weeks, HR 2.52, 1.33–4.78; p = 0.013), and overall survival (median 34 vs 51 weeks; HR 3.29, 1.60–6.74; p = 0.0057), whereas *PIK3CA* exon 9 mutations had no significant effect on response rate, median PFS, and median overall survival.

Taken together, these data highlight the role of *PIK3CA* exon 20 mutations in predicting resistance to cetuximab and panitumumab, although this association should be confirmed in prospective trials. The different impact on clinical outcome exerted by exon 9 and exon 20 mutations is explained by *in vitro* studies, demonstrating that mutations located in different hotspots give rise to different biochemical and oncogenic properties and are differently activated by RAS.²³ Conflicting results from previous published works^{6,22} could therefore be explained by the heterogeneity of patients series in terms of the distribution of mutations in the two different exons.

Pharmacological inhibition of PI3K for cancer treatment is a strategy currently under investigation in several phase I and II trials. Given the frequency and role of oncogenic *PIK3CA* mutations in mCRC above described, it would be rationale to target this pathway in the *KRAS* wild-type population. Indeed, the AKT inhibitor MK-2206 is currently ongoing phase II testing in mCRC chemorefractory patients with *KRAS* wild-type, *PIK3CA*-mutated, mCRC [ClinicalTrials.gov Identifier: NCT01186705] or together with EGFR-targeted monoclonal antibodies to circumvent resistance. In light of recent data,⁴ the latter approach should hypothetically be restricted to the rare subset of patients harboring exon 20 mutations. On the other hand, because of the interaction between RAS and PI3K, ¹⁶ oncogenic *KRAS* itself can prompt cancer cells for escaping pharmacological MEK blockade by activating feedback loop between RAF-MEK-ERK and PI3K pathways.²⁴ Consequently, in a breast cancer preclinical models, dual inhibition with MEK and PI3K inhibitors result in a synergistic tumor growth inhibition.²⁵ Therefore, concomitant PI3K and MEK inhibition appear a promising strategy for *KRAS* mutant-mCRCs, potentially overcoming resistance conferred by compensatory cross-talk between pathways. A phase I clinical trial applying this approach with the PI3K inhibitor BKM120 given in combination with the MEK inhibitor GSK1120212 is currently ongoing in patients with advanced solid tumors selected for *KRAS/BRAF* mutations [ClinicalTrials.gov Identifier: NCT01155453].

Targeting other cell-surface receptors

A different pharmacological strategy to treat *KRAS* mutant tumors is represented by targeting receptors tyrosine kinase other than EGFR that contribute to enhanced cell survival and proliferation. The type 1 insulin-like growth factor receptor (IGF-1R) is a member of a family of transmembrane tyrosine kinases that includes the insulin receptor and the insulin receptor-related receptor. The IGF-1R signaling pathway is an important pathway in different types of cancers including CRC^{26} and include transduction of the IGF signal by the mitogen-activated protein kinase and PI3K/Akt pathways. ²⁷ Recent evidence suggested a role for IGF-1R signaling in the acquired resistance to EGFR inhibitors in glioblastoma cells ²⁸ and there is evidence for cross-talk between IGF-1R and EGFR. ²⁹ Basing on these findings and on preclinical data showing that combination treatment of IGF-1R and EGFR kinase inhibitors result in synergy of growth inhibition in CRC cell lines, ²⁹ a phase II study with the anti-IGF-1R monoclonal antibody IMC-A12, either alone or in combination with cetuximab, was performed in patients with cetuximab- or panitumumab-refractory mCRC. In this study, $30\,$ 64 patients were treated (23 patients with IMC-A12 monotherapy, 21 with IMC-A12 plus cetuximab and 20 with IMC-A12 plus cetuximab among patients who had disease control on a prior anti-EGFR monoclonal antibody and wild-type *KRAS* tumors). No antitumor activity was seen in the 23 patients treated with IMC-A12 monotherapy and of the 21 patients treated with the combination with cetuximab, one patient (with *KRAS* wild-type) achieved a partial response, with disease control lasting 6.5 months. No additional antitumor activity was observed in patients treated with IMC-A12 plus cetuximab who showed disease control on a prior anti-EGFR monoclonal antibody and wild-type *KRAS* tumors. These results indicate no meaningful antitumor activity in this setting (overall, 1 response out of 64 patients) and do not suggest further development of the drug in this setting. Nevertheless, it is interesting to note that the one patient responding to the

combination of IMC-A12 and cetuximab had a tumor that was wildtype for *KRAS*, *NRAS*, *BRAF*, and *PIK3CA.* The authors concluded that *KRAS* wild-type status may be required (but not sufficient) to confer IGF-1R dependence, thus suggesting that this approach is not appropriate for *KRAS* mutant mCRC. Further studies with anti-IGF-1R agents are ongoing in mCRC, including a phase II biomarker study led at our Institutions with the anti-IGF-1R monoclonal antibody AMG 479. In this study (ClinicalTrials.gov Identifier: NCT00891930), mCRC *KRAS* wild-type patients pretreated with irinotecan- and oxaliplatin- or oxaliplatin-based chemotherapy undergo a baseline tumor biopsy and then receive panitumumab with irinotecan (part 1 of the study); at disease progression patients that have displayed response or stable disease undergo a second tumor biopsy and then proceed to part 2 of the study including treatment with panitumumab in combination with AMG 479 with the aim of overcoming acquired resistance to EGFR-targeted therapy. This study will provide insights about mechanisms of secondary resistance (i.e. potential change in *KRAS* mutation status from wild-type at baseline to mutant at the time of the second biopsy following evidence of acquired resistance to panitumumab and irinotecan) and about the potential role of IGF-1R-targeted therapy in overcoming resistance to panitumumab.

The hepatocyte-growth factor (HGF)-mesenchymal epithelial transition factor (MET) molecular pathway is also well known as an important pathway in cancer development. Moreover, METrelated signal transduction is thought to be involved in the development of resistance to EGFR targeting agents³¹ and the combinatorial inhibition of HGF-MET and EGFR is therefore an interesting approach to assess in clinical trials. ³²

In conclusion, RAS plays a central role in EGFR and other receptors tyrosine kinase signaling, thus its constitutive activation could also hamper approaches involving inhibition of IGF-1R and MET pathway. Therefore, in *KRAS* mutant CRC patients, a multi-targeted strategy including combination of both MET or IGF-1R inhibitors together with inhibitors of targets downstream of RAS is probably the best approach. Interestingly, concomitant blockade of IGF-1R and MEK has been shown effective to prevent the occurrence of the EGFR-IGF1R cross-talk and showed preclinical activity in BRAF mutated CRC preclinical models. ³³ Therefore, combinatorial clinical studies might be warranted for chemorefractory mutated mCRC.

Conflict of interests

All authors have no conflict of interest to declare.

Funding

This work was supported by grants from Italian Association for Cancer Research (AIRC) and OCGO (Oncologia Ca' Granda Onlus) Fondazione.

References

- 1. Winer E, Gralow J, Diller L, et al. American Society of Clinical Oncology. Clinical cancer advances 2008: major research advances in cancer treatment, prevention, and screening - a report from the American Society of Clinical Oncology. J Clin Oncol 2008;27:812–26.
- 2. Bardelli A, Siena S. Molecular mechanisms of resistance to cetuximab and panitumumab in colorectal cancer. J Clin Oncol 2010;28:1254–61.
- 3. Allegra CJ, Jessup JM, Somerfield MR, et al. American Society of Clinical Oncology provisional clinical opinion: testing for KRAS gene mutations in patients with metastatic colorectal carcinoma to predict response to anti-epidermal growth factor receptor monoclonal antibody therapy. J Clin Oncol 2009;27:2091–6.
- 4. De Roock W, Claes B, Bernasconi D, et al. Effects of KRAS, BRAF, NRAS, and PIK3CA mutations on the efficacy of cetuximab plus chemotherapy in chemotherapyrefractory metastatic colorectal cancer: a retrospective consortium analysis. Lancet Oncol 2010;8:753–62.
- 5. Di Nicolantonio F, Martini M, Molinari F, et al. Wildtype BRAF is required for response to panitumumab or cetuximab in metastatic colorectal cancer. J Clin Oncol 2008;26:5705–12.
- 6. Sartore-Bianchi A, Martini M, Molinari F, et al. PIK3CA mutations in colorectal cancer are associated with clinical resistance to EGFR-targeted monoclonal antibodies. Cancer Res 2009;69:1851–7.
- 7. Sartore-Bianchi A, Bencardino K, Di Nicolantonio F, et al. Integrated molecular dissection of the epidermal growth factor receptor (EGFR) oncogenic pathway to predict response to EGFR-targeted monoclonal antibodies in metastatic colorectal cancer. Target Oncol 2010;1:19–28.
- 8. Sridhar SS, Hedley D, Siu LL. Raf kinase as a target for anticancer therapeutics. Mol Cancer Ther 2005;4:677–85.
- 9. Rajagopalan H, Bardelli A, Lengauer C, et al. Tumorigenesis: RAF/RAS oncogenes and mismatchrepair status. Nature 2002;418:934.
- 10. Italiano A, Hostein I, Soubeyran I, et al. KRAS and BRAF mutational status in primary colorectal tumors and related metastatic sites: biological and clinical implications. Ann Surg Oncol 2010;17:1429–34.
- 11. Loupakis F, Ruzzo A, Cremolini C, et al. KRAS codon 61, 146 and BRAF mutations predict resistance to cetuximab plus irinotecan in KRAS codon 12 and 13 wildtype metastatic colorectal cancer. Br J Cancer 2009;18:715–21.
- 12. Wilhelm SM, Carter C, Tang L, et al. BAY 43-9006 exhibits broad spectrum oral antitumor activity and targets the RAF/MEK/ERK pathway and receptor tyrosine kinases involved in tumor progression and angiogenesis. Cancer Res 2004;64: 7099–109.
- 13. Kopetz S, Desai J, Chan E, et al. PLX4032 in metastatic colorectal cancer patients with mutant BRAF tumors. J Clin Oncol 2010;28(suppl):15s, Abstr. 3534.
- 14. Hatzivassiliou G, Song K, Yen I, et al. RAF inhibitors prime wild-type RAF to activate the MAPK pathway and enhance growth. Nature 2010;464:358–9.
- 15. Poulikakos PI, Zhang C, Bollag G, Shokat KM, Rosen N. RAF inhibitors transactivate RAF dimers and ERK signalling in cells with wild-type BRAF. Nature 2010;464:427–30.
- 16. Yarden Y. The EGFR family and its ligands in human cancer. Signalling mechanisms and therapeutic opportunities. Eur J Cancer 2001;37(Suppl 4): S3–8.
- 17. Velho S, Oliveira C, Ferreira A, et al. The prevalence of PIK3CA mutations in gastric and colon cancer. Eur J Cancer 2005:41:1649–54.
- 18. Sartore-Bianchi A, Di Nicolantonio F, Nichelatti M, et al. Multi-determinants analysis of molecular alterations for predicting clinical benefit to EGFR-targeted monoclonal antibodies in colorectal cancer. PLoS One 2009;4:e7287.
- 19. Siena S, Sartore-Bianchi A, Di Nicolantonio F, Balfour J, Bardelli A. Biomarkers predicting clinical outcome of epidermal growth factor receptor-targeted therapy in metastatic colorectal cancer. J Natl Cancer Inst 2009;101:1308–24.
- 20. Jhawer M, Goel S, Wilson AJ, et al. PIK3CA mutation/PTEN expression status predicts response of colon cancer cells to the epidermal growth factor receptor inhibitor cetuximab. Cancer Res 2008;68:1953–61.
- 21. Souglakos J, Philips J, Wang R, et al. Prognostic and predictive value of common mutations for treatment response and survival in patients with metastatic colorectal cancer. Br J Cancer 2009;101:465–72.
- 22. Prenen H, De Schutter J, Jacobs B, et al. PIK3CA mutations are not a major determinant of resistance to the epidermal growth factor receptor inhibitor cetuximab in metastatic colorectal cancer. Clin Cancer Res 2009;15:3184–8.
- 23. Zhao L, Vogt PK. Helical domain and kinase domain mutations in p110alpha of phosphatidylinositol 3-kinase induce gain of function by different mechanisms. Proc Natl Acad Sci USA 2008;105:2652–7.
- 24. Yu CF, Liu ZX, Cantley LG. ERK negatively regulates the epidermal growth factormediated interaction of Gab1 and the phosphatidylinositol 3-kinase. J Biol Chem 2002;277:19382–8.
- 25. Mirzoeva OK, Das D, Heiser LM, et al. Basal subtype and MAPK/ERK kinase (MEK)-phosphoinositide 3-kinase feedback signaling determine susceptibility of breast cancer cells to MEK inhibition. Cancer Res 2009;69:565–72.
- 26. Reinmuth N, Liu W, Fan F, et al. Blockade of insulin-like growth factor I receptor function inhibits growth and angiogenesis of colon cancer. Clin Cancer Res 2002; 8:3259–69.
- 27. Wang Y, Sun Y. Insulin-like growth factor receptor-1 as an anti-cancer target: Blocking transformation and inducing apoptosis. Curr Cancer Drug Targets 2002; 2:191–207.
- 28. Chakravarti A, Loeffler JS, Dyson NJ. Insulin-like growth factor receptor I mediates resistance to anti-epidermal growth factor receptor therapy in primary human glioblastoma cells through continued activation of phosphoinositide 3-kinase signaling. Cancer Res 2002;62:200–7.
- 29. Hu YP, Patil SB, Panasiewicz M, et al. Heterogeneity of receptor function in colon carcinoma cells determined by cross-talk between type I insulinlike growth factor receptor and epidermal growth factor receptor. Cancer Res 2008;68:8004–13.
- 30. Reidy DL, Vakiani E, Fakih MG, et al. Randomized, phase II study of the insulinlike growth factor-1 receptor inhibitor IMC-A12, with or without cetuximab, in patients with cetuximab- or panitumumab-refractory metastatic colorectal cancer. J Clin Oncol 2010 Aug 16.
- 31. Stommel JM, Kimmelman AC, Ying H, et al. Coactivation of receptor tyrosine kinases affects the response of tumor cells to targeted therapies. Science 2007; 318:287–90.
- 32. Karamouzis MV, Konstantinopoulos PA, Papavassiliou AG. Targeting MET as a strategy to overcome crosstalk-related resistance to EGFR inhibitors. Lancet Oncol 2009;10:709–17.
- 33. Buck E, Eyzaguirre A, Rosenfeld-Franklin M, et al. Feedback mechanisms promote cooperativity for small molecule inhibitors of epidermal and insulin-like growth factor receptors. Cancer Res 2008;68:8322–32.